Path:
nntp-server.caltech.edu!news.claremont.edu!paris.ics.uci.edu!news.service.uci.edu!ihnp4.ucsd.edu!biosci!bcm!lodi
From: lodi@dino.qci.bioch.bcm.tmc.edu (bill)
Newsgroups: bionet.xtallography
Subject: Re: Crystal Screen cover slips
Date: 21 Jul 1995 21:18:39 GMT
Organization: X-Ray Crystallography / H.H.M.I.
Lines: 27
Distribution: world
Message-ID: <3up5jf$gvh@gazette.bcm.tmc.edu>
References: <806346120-0-18554@blue.weeg.uiowa.edu>
NNTP-Posting-Host: dino.qci.bioch.bcm.tmc.edu

In article <806346120-0-18554@blue.weeg.uiowa.edu>
sfredric@blue.weeg.uiowa.edu writes:
>Thanks to all who responded to my previous query concerning glycerol and
>crystallization.
>
>And now. . . .I would appreciate hearing about efficient ways to siliconize
>coverslips. My 1st few attempts, using Sigmacote, have generated spotty
>coverslips. Thanks, Scott

Tell you what... I'll trade you. For siliconizing cover slips with dichlorodimethylsilane, make a 5% v/v solution in toluene. Have a beaker of toluene ready for the rinse, DON'T use water at this stage, it will make a gummy mess. Rinse with ethanol, then ddH2O. Remember dichlorodimethylsilane is nasty stuff.

Also, this 5% solution can be diluted 1:100 with toluene to give a 0.05% solution which can be used to briefly rinse x-ray capillaries (then rinse with ethanol), yielding a lightly siliconized cap that cleans up easily (i.e. quick to get rid of excess m.l.) without giving crystal slippage. (Your mileage may vary.)

Now, how about a summary of the glycerol tips?

Thanks

W. E. Meador